

Association Between Variants of Neuromedin U Gene and Taste Thresholds and Food Preferences in European Children

Results From the IDEFICS Study

Grippi, Claudio; Ahrens, Wolfgang; Buchecker, Kirsten; Chadjigeorgiou, Charalambos; De Henauw, Stefaan; Koni, Anna C.; Foraita, Ronja; Lissner, Lauren; Molnar, Dénes; Moreno, Luis Alberto; Pitsiladis, Yannis; Reisch, Lucia A.; Russo, Paola; Siani, Alfonso; Veidebaum, Toomas; Iacoviello, Licia; Gianfagna, Francesco

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5 **2 in European children:**
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8 **3 Results from the IDEFICS study**
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94 41 **Declarations of interest:** none

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121 50 **ABSTRACT (max 280 words- *Appetite*)**
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128 53 **Aim:** The neuropeptide neuromedin U (NMU) known for its role in appetite, feeding and energy
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130 54 balance could be involved in the control of food choice and taste sensitivity. We examined the
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132 55 association between *NMU* polymorphisms/haplotypes and taste thresholds and food preferences in a
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134 56 population of European children.

135
136 57 **Methods:** A total of 578 subjects from the IDEFICS study (mean age 7.5±0.8 SD, boys 53.6%)
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138 58 with *NMU* genotype data and food preference (salty, fatty, sweet, flavour and umami food) and
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140 59 taste threshold (salt, fat, sweet, umami) tests available were analysed. Three single nucleotide
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142 60 polymorphisms (SNPs; rs6827359, T:C; rs12500837, T:C; rs9999653, C:T) of *NMU* gene were
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144 61 analyzed and five major haplotypes were inferred. The associations between genotypes and food
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146 62 preferences or taste thresholds were investigated (odds ratios –OR, adjusted for age, sex and
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148 63 country). A $p < 0.05$ after false discovery rate adjustment (p FDR) was considered statistically
149
150 64 significant.

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153
154 65 **Results:** The association between *NMU* genotypes and food preference showed two *NMU* SNPs
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156 66 associated with preference for food containing sodium glutamate (umami taste; rs6827359C,
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158 67 OR=1.61, 95% confidence interval (CI):1.20-2.17; rs9999653T, OR=1.59, 95%CI:1.18-2.13). In the
159
160 68 haplotype analysis, the CTT haplotype showed an OR of 1.70 (95%CI:1.16-2.5) for the umami food
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162 69 preference, while CCT haplotype showed an OR of 1.63 (95%CI:1.11-2.40), compared to the most
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164 70 frequent haplotype (TTC). Carriers of CCT/CCT vs subjects with no CCT haplotype showed an OR
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166 71 of 4.78 (95%CI:1.86-12.30). Umami food preference was associated with low values of BMI z -
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168 72 score, arm circumferences, skinfolds and fat mass (p FDR<0.05). No association between *NMU*
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170 73 genetic variants and taste thresholds was found.
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74 **Conclusions:** This study shows for the first time in children an association between preference for
75 umami food and a *NMU* haplotype, previously found associated with low BMI values.

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77 **Keywords:** *food preferences; umami; neuromedin U; neurology; genetics; obesity*

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239 84 **INTRODUCTION**
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242 85 Taste sensing influences food preference, appetite and satiety, thereby regulating diet quality
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244 86 and total food intake and, as a result, weight maintenance (Dotson 2012). Five taste qualities can be
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246 87 perceived by humans: sweet, salty, bitter, sour and umami. The ability of discriminate between
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248 88 tastes has shown to be the result of evolution, to avoid hazardous compounds while searching for
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250 89 nutrients important for life and development.
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252
253 90 In children, food preferences are often guided by taste alone. Specifically, preferences for
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255 91 sugar and fat may be acquired early in life, as children learn to prefer those flavors that are
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257 92 associated with high energy density and fat content, with higher risk of developing overweight
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259 93 (Drewnowski 1997). In line with that, several studies showed that food preferences and taste
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261 94 sensitivity differ between obese and non-obese children (Overberg 2012; Wardle 2001).
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265 95 The mechanisms behind the regulation of food preferences and taste perception has only
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267 96 been partially revealed (MacLean 2017). Recent findings suggested that genetics plays an important
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269 97 role, since high heritability levels were found for both food preferences and taste perception
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271 98 (Tornwall 2015). Genetic studies of taste variability have focused on a number of candidate genes
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273 99 encoding the taste receptors, hormones and neuropeptides, such as leptin, GLP-1 and NPY,
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275 100 important modulators at both peripheral and central level (Feeney 2011; Loper 2015).
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278 101 Neuromedin U (NMU) is a neuropeptide with a highly conserved genetic structure, thought
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280 102 to have several important functions. Transgenic mouse models and experimental studies showed its
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282 103 main involvement in the regulation of body weight, through its effect on appetite, feeding and
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284 104 energy balance (Martinez 2015). Recently, an increased preference for obesogenic food was
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286 105 observed in rats knockdown for NMU Receptor 2 (NMUR2), the NMU receptor mainly expressed
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288 106 in the central nervous system (Benzon 2014). Although the effects of NMU are well understood in
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290 107 animal models, little is known about its role in humans besides a suggested role in adiposity
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108 regulation from epidemiological evidence. A rare NMU variant was in fact associated with
109 overweight and obesity (Hainerová 2006). In addition, our group observed an association between
110 NMU genetic polymorphisms and adiposity indices in a European children population (Gianfagna
2017). While a link between NMU and adiposity regulation was identified, no data have been
produced to link this neuropeptide to adiposity intermediate phenotypes. To this respect, NMU
could cooperate in modifying taste perception and selective appetite.

In this study, we aimed at evaluating the potential association of NMU with food preferences and taste perception, by investigating NMU genetic variants in a European children population recruited for the IDEFICS Study (Ahrens 2011).

MATERIALS AND METHODS

Study population

IDEFICS (Identification and prevention of dietary - and lifestyle - induced health effects in children and infants) is a large European multi-center study aimed at investigating risk factors associated with childhood obesity (Ahrens 2011). A cohort of 16,229 children aged 2.0–9.9 years was recruited in a population-based survey between September 2007 and May 2008 (T0), in eight European countries (Belgium, Cyprus, Estonia, Germany, Hungary, Italy, Spain and Sweden).

For taste threshold and food preference test, from the subgroup of participating primary schoolchildren aged 6–9 or 7–9 years (depending on age of school enrolment), a subsample of 1839 (20.8%) children were randomly selected (Lanfer 2012). For genetic analyses, a subgroup of 4,678 (28.8%) samples was randomly selected from the total study population of European descent

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357 133 children, stratifying by age, sex and country (about 600 subjects from each country) (Gianfagna
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359 134 2013; Cugino 2013). Finally, 578 children, 6–9-year-old with *NMU* genotype and food preference
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361 and taste threshold test available were selected for the present analysis.
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364 136 Ethical approval was obtained by the ethical committees of each center engaged in the
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366 137 fieldwork: Ethics Committee, University Hospital, Gent, Belgium; Cyprus National Bioethics
367
368 138 Committee, Strovolos, Cyprus; Tallinn Medical Research Ethics Committee, Tallinn, Estonia;
369
370 139 Ethics Committee, University of Bremen, Bremen, Germany; Egészségügyi Tudományos Tanács,
371
372 140 Pécs, Hungary; Comitato Etico, ASL Avellino, Avellino, Italy; Comité Ético de Investigación,
373
374 141 Clínica de Aragón (CEICA), Zaragoza, Spain; Regional Ethics Committee, University of
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376 142 Gothenburg, Gothenburg, Sweden. Both children and their parents gave oral (children) and written
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378 (parents) informed consent.
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382 383 145 **Data collection**

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385 146 *Food preference test.* The preference test was organized as paired and forced choice on a
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387 147 board as previously described (Lanfer 2012, Knof 2011). Briefly, participating children had their
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389 148 last meal 1 h before to ensure that they were neither hungry nor sated. The test was conducted using
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391 149 five preference tests for five tastes. To evaluate sweet preference apple, juice was administered in
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393 150 small cups with a volume of 30 ml at 18 ± 2 °C and with different addition of sucrose (0.53-3.11%).
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395 151 For the evaluation of flavour preferences, 0.05% apple flavour (nature identical, Sensient Flavors,
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397 Bremen, Germany) was added to the basic recipe. The children had to rinse their mouths with water
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399
400 153 between each pair sequence of the test. To assess the preference for salty, fatty and umami tastes,
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402 154 crackers were selected as food sample. The crackers were covered with 0.5% aqueous solution of
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404 155 sodium hydroxide (soda lye, Carl Roth Chemicals, Karlsruhe, Germany) to make them tastier to the
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406 156 children. The basic recipe of cracker included water, flour (wheat), fat (8%) and salt. The same type
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408 157 of crackers was modified with an increased 8% of fat to assess high-fat preference, an increased 1%
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410 158 of salt for salty taste and monosodium glutamate (1%) for umami crackers. In each sequence, the
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416 159 children had to choose their preferred food sample between the basic recipe and a modified one.
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418 160 The order of assessment for the food choice was fat, salt and umami. The test procedure was subject
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421 161 to pre-testing before the beginning of the study (Suling 2011) and yielded reliable results in a
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423 162 reproducibility study (Knof 2011).

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425 163 ***Taste detection threshold test.*** As a measure of taste sensitivity, detection threshold is the
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427 164 lowest value of a tastant that must be exceeded to have any effect on the observer. The procedure to
428
429 165 evaluate the taste threshold was described in detail in Knof et al. (2011). In brief, a paired
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431 166 comparison test with five different watery solutions at different concentrations of tastants was
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433 167 served to the children in small cups of 20 ml. The concentration ranges of the tastants were sucrose
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435 168 (8.8-46.7 mmol/l), sodium chloride (3.4-27.4 mmol/l), caffeine (0.26-1.3 mmol/l) and monosodium
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438 169 glutamate (MSG) (0.6-9.5 mmol/l). The paired test was prepared as a board game and the children
439
440 170 had to compare each test solution at increasing concentrations of tastants with a cup containing pure
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442 171 water, to find the cups that would taste differently from the previous one. Taste detection at lower
443
444 172 tastant concentration (lower detection threshold value) indicates increased sensitivity for a specific
445
446 173 tastant. Between the taste modalities, the children had to neutralize their taste with distilled water.

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448 174 ***Anthropometric data.*** Height was measured using a standard clinical Seca 225 stadiometer
449
450 175 (Seca, Hamburg, Germany) to the nearest 0.1 cm, and weight was measured using a scale (BC 420
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452 176 SMA; Tanita, Amsterdam, The Netherlands) to the nearest 0.1 kg, on children wearing underwear
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454
455 177 clothes and without shoes. BMI was calculated as $\text{weight(kg)/height(m)}^2$. Waist and hip
456
457 178 circumference was measured with an inelastic tape (Seca 200, precision 0.1 cm, range 0–150 cm).
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459 179 Age- and sex-specific BMI and waist circumference z-scores and BMI categories were calculated
460
461 180 according to the criteria of the International Obesity Task Force (IOTF) (Cole 2012). Leg-to-leg
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463 181 impedance was measured with the Tanita scale and fat-free mass was calculated using the formula
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465 182 of Tyrrell *et al* (2001). Skinfold thicknesses (tricipital and subscapular) were measured with a
466
467 183 Holtain caliper (Holtain, Holtain Ltd, Pembrokeshire, UK, range 0±40 mm), taking measures twice
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469 184 on the right hand body side and using the mean of the two measures for the analyses. All
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475 185 measurements were collected by standardized protocols across centers, checking intra- and inter-
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477 186 observer reliability (Stomfai 2011).

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480 187 **Genotyping.** DNA extraction was carried out from saliva samples (Oragene DNA Self-
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482 188 Collection Kit, OG-300/OG-250; DNA Genotek Inc., Kanata, Ontario, Canada) (Koni 2011).
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484 189 Among the three main blocks of the *NMU* gene (chr4, 55595229–55636698, GRCh38.p7
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486 190 assembly), three tag SNPs (rs6827359, rs12500837, rs9999653; intronic regions) were selected
487
488 191 from the Caucasian HapMap Project data using the Tagger Pairwise method of Haploview software
489
490 192 (version 4.1; Broad Institute, Cambridge, MA, USA) (Barrett 2005). Tag SNP selection criteria
491
492 193 were described in Gianfagna et al (2017). The SNPs were genotyped by a multiplexed end-point
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494 194 assay. The allelic discrimination was performed by 7500 Fast Real-Time System (Applied
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496 195 Biosystems). The genotyping success rate was on average 97.6% and a randomly selected sample
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498
499 196 (5%) was newly genotyped for all SNPs with 100% concordance.

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503 198 **Statistical analysis**

504
505 199 The analyses were conducted with SAS (v9.3, SAS Institute Inc., Cary, NC) and R (v3.2.1;
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507 200 <https://www.R-project.org/>) software. Distribution of continuous variables was assessed using the
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509 201 Kolmogorov–Smirnov test and log-transformed variables were used where appropriate. Hardy-
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511 202 Weinberg equilibrium (HWE) was assessed with the chi-square test. The best genetic model was
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513 203 checked for each genotype–phenotype association, testing dominance deviation from additivity and
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515 204 considering the additive model as default (Hoffman 2004). The Haplo.stats package of R software
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517 205 was used to estimate the haplotype frequencies and to verify the associations between haplotype and
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519 206 phenotype (haplo.glm function, the most prevalent haplotype as reference). Haplotypes with
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521 207 frequencies lower than 1% were excluded. Multiple regression analyses were performed using age,
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523 208 sex and countries as covariates, to evaluate the association between genotypes and food preferences
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525 209 (basic food or modified food with salty, fatty, umami, sweet and flavor tastes) or taste thresholds
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527 210 (sweet, salty, umami, bitter). The Benjamini-Hochberg false discovery rate (FDR) (Benjamini

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211 1995) was used to adjust the results for multiple comparisons, using PROC MULTTEST in SAS. A
212 FDR-adjusted p value (p_{FDR}) <0.05 was considered as statistically significant.

RESULTS

Population characteristics

Characteristics of the study population are shown in Table 1. Children with at least one SNP successfully genotyped in NMU gene and eating behavior data available were 578 (mean age 7.5 ± 0.8 SD, boys 53.6%). The lowest taste threshold value was found for sweet taste, taking into account mean thresholds and concentration ranges of test solutions (sucrose, 19.9 mmol/l; range 8.8-46.7). In food preference tests, the most appreciated food were salt-added crackers (63.7% of children preferred them to the basic recipe crackers), while sodium glutamate-added crackers (umami taste) were the less appreciated (33.4%). Genotype frequencies are reported in Table 2a. No statistically significant differences were found between genotype frequencies of children randomly selected to be included or excluded from this analysis subjects (data not shown). All genotypes were in Hardy-Weinberg equilibrium and the minor allele frequencies (MAF) were similar to values reported in the HapMap database for Caucasians. Six haplotypes were inferred (Table 2b; wild-type haplotype 43.9%).

Associations with food preferences and taste thresholds

Deviation from additivity test showed codominance as the best genetic model for all phenotype-SNP and phenotype-haplotype associations, except for umami food preference for the reference haplotype TTC (H7), that showed a dominant model.

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592
593 235 The analysis of the association between *NMU* genetic variants and food preferences are
594 shown in Table 3. Children carrying mutant allele in Two-two out of the three *NMU* SNPs studied
595 236 were more likely to prefer associated with preference for food containing sodium glutamate (umami
596 taste) with respect to the same food prepared with a basic recipe (;rs6827359, odds ratio -
597 OR=1.61, 95% confidence interval - 95%CI:1.20-2.17; rs9999653, OR=1.59, 95%CI:1.18-2.13;
598 237 Table 3). The haplotype analysis confirmed the association between genetic variants and preference
599 for with umami preference food. Subjects carrying the CTT haplotype showed an OR of 1.70
600 238 (95%CI:1.16-2.50) for the umami preference, while CCT haplotype showed an OR of 1.63
601 (95%CI:1.11-2.40), as compared to the most frequent TTC haplotype. The reference haplotype
602 239 showed a reduced odd for umami preference when compared to all other haplotypes (OR=0.42;
603 95%CI:0.27-0.66). The association between umami food preference and the two SNPs as well as
604 240 the TTC haplotype remained statistically significant after FDR correction. The same analysis
605 considering each single genotype or haplotype versus the correspondent wild-type (Table 3, last
606 241 rows) showed even higher association values. Homozygotes for mutant allele of rs6827359
607 (OR=2.57; 95%CI:1.43-4.61) and of rs9999653 (OR=2.31; 95%CI:1.29-4.15) versus the opposite
608 242 homozygotes showed FDR-significant association with umami food preference. Similarly,
609 homozygotes for CTT (OR=4.78; 95%CI:1.86-12.30), as well as carriers of TTC (OR=0.43;
610 243 95%CI:0.26-0.68) haplotype showed FDR-significant association with umami food preference
611 when compared with subjects without that specific haplotype. No significant associations were
612 244 found between *NMU* genotypes or haplotypes and the other food preferences (Suppl. Table 1).
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635 The analysis of the association between *NMU* genetic variants and taste thresholds showed
636 255 no significant results (Table 4, codominant model, and Suppl. Table 2, genotype or haplotype
637 contrasts).
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642 258 643 644 259 **Association between umami preference and adiposity indices** 645 646 647 648 649

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652 260 Preference for umami food was associated with lower levels of the main anthropometric
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654 261 measures (Table 5, Model 1), such as *z*-score BMI (-0.25, *p*=0.034) and fat mass (-0.93, *p*=0.012).
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656 262 The association between preference for umami food and overweight/obesity was not statistically
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659 263 significant (OR=0.70; 95%CI:0.42-1.17). An association was also found with skinfolds (sum of
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661 264 triceps and subscapular skinfold, -1.75, *p*=0.025) and arm circumference (-0.63, *p*=0.011). The
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663 265 association between umami and anthropometric measures did not change when NMU genetic
664
665 266 variants were added as covariates in the regression model (Table 5, Model 2). The association
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667 267 between the NMU haplotypes and anthropometric measures, previously observed in the whole
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669 268 sample of subjects with genetic data available, was not significant in this smaller subsample of
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671 269 children with taste thresholds and food preference data.
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673 270 674 675 676 271 677 678 272 **DISCUSSION**

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682 274 In this study, conducted in a sub-sample of children from the large European IDEFICS
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684 275 cohort, we found variants (two SNPs and two haplotypes) of *NMU* gene associated with preference
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686 276 for food containing glutamate. Preference for umami food was inversely associated with several
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688 277 anthropometric parameters such as BMI, weight, waist and arm circumference, skinfolds and fat
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690 278 mass.

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692 279 This is the first study in humans investigating the potential involvement of NMU in
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695 280 influencing food preferences and taste perception in humans, although some experimental studies
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697 281 suggested this potential role. A recent experimental study in rats revealed that NMUR2, one of the
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699 282 two NMU receptors, could induce preference for high-fat foods (Benzon 2014). The NMUR2 is
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701 283 located mainly in central nervous system and is highly expressed in the hypothalamous and in other
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703 284 regions receiving fibers from taste sensory ganglia (Stanska 2016; Li 2017). Our results on the
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705 285 association between NMU genetic variants and umami food preference support the role of NMU in
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711 286 regulating feeding behavior (Jethwa 2005; Kowalski 2005), specifically in neuronal pathways of
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713 287 taste-like/dislike preferences for umami food. We found no association with other food preferences.
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716 288 In line with our negative findings, a recent experimental study in mice showed that a NMU analog
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718 289 has no effect on preference for a sweet drink (Kaisho 2017). Although several hormones and
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720 290 neuropeptides were suggested as modulators of peripheral gustatory system (Loper 2015), we found
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722 291 no association between *NMU* genetic variants and taste perception thresholds. The reason of this
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724 292 negative result could be a lack of expression of NMU receptors in oral cavity. In fact, we found no
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726 293 literature data on expression of NMUR1 and NMUR2 in oral cavity, although at least NMUR1 is
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728 294 abundantly present along the gastrointestinal tract (Li 2017; Hedrick 2000; Raddatz 2000). These
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730 295 findings confirm that the associations of NMU with food preference is mediated by central nervous
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733 296 and not peripheral gustatory system.

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735 297 In our population, we found significant associations between umami preference and
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737 298 anthropometric parameters. Although some associations did not reach statistical significance, all
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739 299 were concordantly in the direction of a protective effect for umami preference. The strongest
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741 300 significant association was found for fat mass. Children with preference for umami food present a
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743 301 decreased fat mass of approximately 1 kg. The umami taste is the fifth taste identified (Ikeda 2002),
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745 302 induced principally by three molecules: monosodium glutamate (MSG), inosine-5'-monophosphate
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747 303 (IMP) and guanylo-5'-monophosphate (GMP). These substances enhance the savory of foods and
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749 304 cause sensation such as pleasure and satisfaction (De Araujo 2003; Kurihara 2015), as well as
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752 305 insulin release, salivary, gastric and pancreatic secretion, gastric emptying and distal colon
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754 306 peristalsis (Stanska 2016). These effects could result in the modulation of short-term intake and
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756 307 satiety by giving umami an important role in appetite control (Masic 2014). The variants found
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758 308 associated with an increased preference for umami food were the same identified associated with
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760 309 lower BMI values, in a larger sample of the same population (Gianfagna 2017), suggesting that an
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762 310 increase in food preferences for umami could be also associated with decreased BMI. Supporting
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764 311 this hypothesis, a study in rats showed an association between preference of monosodium glutamate

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770 312 solution and reduction of obesity (Kondoh 2008). This may suggest a potential mediating effect of
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772 313 umami food preference in the association between *NMU* genetic variants and BMI. However, in the
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774 314 subgroup with food preference data available, the association between *NMU* variants and BMI was
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777 315 not significant as we previously reported in the whole sample with genetic data available
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779 316 (Gianfagna 2013) due to the smaller sample size. The regression coefficient was however similar.

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781 317 The strength of this study is the availability of a children population, which represent a good
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783 318 model for genetic studies. In fact, since the environment has had less time to exert its effect,
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785 319 phenotypes have a larger genetic component than in adults. However, taste and food preferences are
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787 320 influenced by several non-genetic factors, such as social, community and environmental factors,
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789 321 operating at multiple levels throughout life (Beckerman 2017; Russell 2013), which were not
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791 322 considered in this analysis. Although a confounding effect should be excluded due to the genetic
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793 323 study design, we cannot exclude a modification of the genetic effect due to factors not considered in
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795 324 the analyses. A further limitation of this study is the sample size, which is underpowered for the
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797 325 mediation effect analysis of umami preferences.

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800 326 In conclusion, variants in the *NMU* gene might play a role in determining umami food
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802 327 preferences. This mechanism could mediate part of the association between the same *NMU* genetic
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804 328 variants and BMI, previously observed, although further studies are necessary to confirm this
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806 329 hypothesis.

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Table 1. Anthropometric characteristics of N = 578 children with genotype and food preferences or taste threshold data available.

| Variables | N | Mean±SD, n (%) |
|---|----------|---------------------------|
| Age (years; mean±SD) | 578 | 7.5 ±0.8 |
| Males (n, %) | 578 | 310 (53.6%) |
| Body Mass Index (mean±SD) | 578 | 16.8 ±2.7 |
| BMI z-score (mean±SD) | 578 | 0.47 ±1.17 |
| Weight (kg; mean±SD) | 578 | 27.3 ±6.1 |
| Waist circumference (cm; mean±SD) | 578 | 57.0 ±7.0 |
| Hip Circumference (cm; mean±SD) | 577 | 67.1 ±6.7 |
| Waist to hip (mean±SD) | 577 | 0.85 ±0.05 |
| Arm circumference (cm; mean±SD) | 572 | 19.7 ±2.5 |
| Skinfolds (sum of tricipital and subscapular, mm; mean±SD) | 570 | 19.1 ±8.1 |
| Fat mass (kg; mean±SD) | 569 | 8.1 ±3.8 |
| Overweight+obese | 578 | 135 (23.4%) |
| Obese | 578 | 49 (8.5%) |
| Children preferring each modified food versus basic recipe | | |
| Fat (n,%) | 460 | 258 (56.1%) |
| Salt (n,%) | 466 | 297 (63.7%) |
| Umami (n,%) | 461 | 154 (33.4%) |
| Sweet (n,%) | 514 | 300 (58.4%) |
| Flavour (n,%) | 518 | 304 (58.7%) |
| Taste detection thresholds | | |
| Sweet (sucrose 8.8–46.7 mmol/l; mean±SD) | 517 | 19.9 ±11.1 |
| Salty (sodium chloride 3.4–27.4 mmol/l; mean±SD) | 522 | 12.9 ±7.2 |
| Bitter (caffeine 0.26–1.3 mmol/l; mean±SD) | 521 | 0.88 ±0.49 |
| Umami (MSG 0.6–9.5 mmol/l; mean±SD) | 516 | 4.2 ±3.1 |

Table 2a. Allele frequencies and Hardy-Weinberg equilibrium of the Neuromedin U (NMU) single nucleotide polymorphisms (SNPs; n = 578 with at least one SNP successfully genotyped).

| SNP | Major:minor allele | N | Homozygous (major allele) | Heterozygous | Homozygous (minor allele) | HWE* (p) | MAF* (%) | CEU (%) |
|------------|--------------------|-----|---------------------------|--------------|---------------------------|----------|----------|---------|
| rs6827359 | T*:C | 576 | 161 (27.9%) | 268 (46.5%) | 147(25.5%) | 0.11 | 49 | 40 |
| rs12500837 | T*:C | 577 | 323 (56.0%) | 216(37.4%) | 38 (6.6%) | 0.91 | 24 | 21 |
| rs9999653 | C:T* | 574 | 132 (23.0%) | 264 (46.0%) | 178(31.4%) | 0.42 | 54 | 49 |

*Hardy-Weinberg Equilibrium (HWE) and Minor Allele Frequency (MAF) were checked on the whole sample of 4649 subjects genotyped;

rs9999653T allele is considered the minor allele based on population frequencies; CEU=Utah Residents (Caucasians) with Northern and Western European Ancestry

Table 2b. Haplotype frequencies (n = 578 with at least one SNP successfully genotyped).

| rs6827359 | rs12500837 | rs9999653 | Freq (%) |
|-----------|------------|-----------|----------|
| T | T | C | 43.2% |
| T | T | T | 7.9% |
| C | C | C | 2.7% |
| C | C | T | 22.6% |
| C | T | T | 23.6% |

Table 3. Odds ratios (OR) for preferring specific modified food to basic recipe, in children carrying genetic variants versus wild-type (codominant model, OR per minor allele).

| Food preferences | Genotypes OR (95%CI) | | | Haplotypes OR (95%CI) | | | |
|--------------------------------|--------------------------|-------------------------|--------------------------|--------------------------|---------------------------|-------------------------|---------------------------|
| | rs6827359 (C) | rs12500837 (C) | rs9999653 (T) | H2 (CCT) | H4 (CTT) | H8 (TTT) | H7 (TTC) ^o |
| Fat | 1.04 (0.79-1.36) | 0.91 (0.67-1.25) | 1.07 (0.82-1.40) | 0.98 (0.70-1.38) | 1.10 (0.77-1.56) | 0.94 (0.57-1.57) | 1.00 (0.76-1.31) |
| Salt | 1.07 (0.81-1.40) | 0.98 (0.71-1.35) | 1.11 (0.84-1.46) | 1.06 (0.74-1.51) | 1.15 (0.81-1.64) | 1.04 (0.62-1.73) | 0.93 (0.71-1.22) |
| Sweet | 1.11 (0.87-1.42) | 0.95 (0.71-1.26) | 1.16 (0.91-1.49) | 1.04 (0.75-1.44) | 1.24 (0.90-1.70) | 1.21 (0.77-1.89) | 0.88 (0.68-1.13) |
| Flavour | 0.93 (0.73-1.20) | 0.90 (0.67-1.20) | 0.89 (0.69-1.15) | 0.85 (0.61-1.18) | 0.98 (0.70-1.36) | 0.94 (0.59-1.51) | 1.08 (0.83-1.41) |
| Umami | 1.61 (1.20-2.17)* | 1.34 (0.96-1.87) | 1.59 (1.18-2.13)* | 1.63 (1.11-2.40) | 1.70 (1.16-2.50) | 1.38 (0.78-2.42) | 0.42 (0.27-0.66)*§ |
| <i>W/M vs W/W</i> [†] | <i>1.17 (0.69-1.97)</i> | <i>1.26 (0.80-1.96)</i> | <i>0.94 (0.54-1.64)</i> | <i>1.56 (0.96-2.53)</i> | <i>1.29 (0.78-2.11)</i> | <i>1.50 (0.76-2.95)</i> | 0.42 (0.26-0.68)* |
| <i>M/M vs W/W</i> [†] | 2.57 (1.43-4.61)* | <i>1.98 (0.87-4.50)</i> | 2.31 (1.29-4.15)* | 2.92 (1.17-7.33) | 4.78 (1.86-12.30)* | <i>1.00 (0.18-5.70)</i> | 0.43 (0.24-0.78) |

Bold: nominally significant associations; *significant also after FDR correction, including rare haplotypes (not shown); § results for dominant model ^oall other haplotypes as reference; [†]Genotype or haplotype contrasts: for genotype analysis, heterozygotes or homozygotes for variant alleles (M) vs homozygotes for wild-type allele (W, ref); for haplotype analysis, subjects with 1 (w/M) or two (M/M) copies of a specific haplotypes versus subjects with the most frequent haplotype (TTC). Analyses adjusted for age, sex, country

Table 4. Differences in detection thresholds for specific tastes, in children carrying genetic variants versus wild-type (codominant model, threshold change per minor allele).

| Taste thresholds | Genotypes (beta±SE, <i>p</i>) | | | Haplotypes (beta±SE, <i>p</i>) | | | |
|------------------|-----------------------------------|-------------------|-------------------|------------------------------------|--------------------|-------------------|-----------------------|
| | rs6827359 (C) | rs12500837 (C) | rs9999653 (T) | H2 (CCT) | H4 (CTT) | H8 (TTT) | H7 (TTC) [°] |
| Sweet | -0.34±0.65 (0.60) | -0.03±0.75 (0.97) | -0.90±0.64 (0.16) | -0.59±0.83 (0.48) | -0.79±0.84 (0.35) | -1.57±1.31 (0.23) | 0.74±0.65 (0.26) |
| Salty | 0.37±0.42 (0.38) | 0.59±0.49(0.23) | -0.18±0.42 (0.66) | 0.35±0.54 (0.52) | -0.15±0.55 (0.78) | -1.57±0.84 (0.06) | 0.07±0.42 (0.87) |
| Bitter | -0.02±0.03 (0.51) | 0.02±0.03 (0.63) | -0.02±0.02 (0.52) | -0.004±0.04 (0.92) | -0.03±0.037 (0.44) | 0.06±0.06 (0.33) | 0.001±0.03 (0.97) |
| Umami | -0.08±0.18 (0.66) | 0.12±0.22 (0.59) | -0.14±0.19 (0.46) | 0.002±0.24 (0.99) | -0.24±0.24 (0.32) | -0.08±0.38 (0.83) | 0.10±0.19 (0.61) |

[°]All other haplotypes as reference. Analysis adjusted for age, sex and country. Lower detection threshold values indicate higher sensitivity for a specific tastant.

Table 5. Associations between anthropometric variables and umami or NMU variants in the IDEFICS children population.

| | N | Model 1 | Model 2 | | | |
|---|----------|-----------------------------|----------------------------|-------------------|-------------------|-------------------|
| Anthropometric | N | Umami preferences | Umami preferences | H2/H2 | H4/H4 | H8/H8 |
| BMI z-score (beta±SE, <i>p</i>) | 456 | -0.25±0.12 (0.034)*° | -0.23±0.12 (0.043)° | -0.12±0.26 (0.64) | -0.15±0.23 (0.52) | 0.22±0.60 (0.71) |
| Waist z-score (beta±SE, <i>p</i>) | 456 | -0.16±0.19 (0.42) | -0.12±0.19 (0.53) | -0.11±0.28 (0.69) | 0.29±0.29 (0.32) | 0.03±0.82 (0.97) |
| Arm circumference (beta±SE, <i>p</i>) | 450 | -0.63±0.25 (0.011)* | -0.58±0.23 (0.011)* | -0.26±0.52 (0.61) | -0.41±0.44 (0.36) | 0.61±1.34 (0.65) |
| Skinfolds [§] (beta±SE, <i>p</i>) | 450 | -1.75±0.78 (0.025)* | -1.71±0.68 (0.012)* | -0.53±1.49 (0.72) | 0.05±1.45 (0.98) | -2.18±1.92 (0.26) |
| Fat mass (beta±SE, <i>p</i>) | 453 | -0.93±0.37 (0.012)* | -0.85±0.34 (0.012)* | -0.21±0.77 (0.78) | -0.74±0.61 (0.23) | 0.98±2.08 (0.64) |
| Overweight/obesity (OR, 95%CI) | 456 | 0.70 (0.42-1.17) | 0.72 (0.44-1.19) | 1.09 (0.40-2.98) | 0.80 (0.29-2.16) | 0.96 (0.09-10.66) |

Model 1: age, sex and country; model 2: all in model 1 plus haplotypes. Bold: nominally significant associations; *significant also after FDR correction; [§] sum of tricipital and subscapular skinfolds; [°] This is equal to -6.1±3.0 (beta±SE) BMI percentiles of the European population distribution (extended definition, Cole et al. 2012) Analysis adjusted for age, sex, country

Supplementary Table 1. Associations between Neuromedin U genotypes and food preferences in the IDEFICS children population.

| Food preferences | Genotype or haplotype contrast† | Genotypes OR (95%CI) | | | Haplotypes OR (95%CI) | | | |
|------------------|---------------------------------|--------------------------|------------------|--------------------------|--------------------------|---------------------------|-------------------|--------------------------|
| | | rs6827359 (C) | rs12500837 (C) | rs9999653 (T) | H2 (CCT) | H4 (CTT) | H8 (TTT) | H7 (TTC)° |
| Fat | W/M vsW/W | 1.06 (0.67-1.68) | 0.83 (0.55-1.25) | 0.84 (0.55-1.25) | 0.91 (0.59-1.40) | 1.04 (0.67-1.61) | 0.81 (0.44-1.49) | 0.88 (0.57-1.36) |
| | M/M vsW/W | 1.07 (0.63-1.84) | 0.98 (0.44-2.19) | 1.13 (0.66-1.92) | 1.15 (0.50-2.67) | 1.35 (0.52-3.49) | 1.84 (0.34-9.97) | 1.03 (0.59-1.79) |
| Salt | W/M vsW/W | 1.21 (0.76-1.93) | 1.05 (0.69-1.60) | 1.22 (0.74-2.02) | 1.14 (0.73-1.80) | 1.23 (0.78-1.94) | 0.87 (0.49-1.56) | 0.97 (0.62-1.54) |
| | M/M vsW/W | 1.12 (0.65-1.94) | 0.85 (0.39-1.88) | 1.24 (0.72-2.13) | 0.96 (0.40-2.30) | 1.15 (0.50-2.65) | 2.99 (0.28-32.52) | 0.85 (0.49-1.47) |
| Sweet | W/M vsW/W | 1.06 (0.69-1.63) | 1.00 (0.68-1.46) | 1.11 (0.70-1.77) | 1.03 (0.69-1.54) | 1.21 (0.81-1.82) | 1.13 (0.66-1.95) | 0.84 (0.56-1.25) |
| | M/M vsW/W | 1.24 (0.75-2.04) | 0.82 (0.40-1.68) | 1.34 (0.81-2.20) | 1.11 (0.48-2.57) | 1.59 (0.71-3.56) | 1.87 (0.47-7.41) | 0.79 (0.47-1.32) |
| Flavour | W/M vsW/W | 1.26 (0.82-1.96) | 0.87 (0.59-1.29) | 1.20 (0.74-1.93) | 0.77 (0.51-1.16) | 0.93 (0.61-1.42) | 0.94 (0.56-1.59) | 1.42 (0.94-2.13) |
| | M/M vsW/W | 0.86 (0.52-1.42) | 0.84 (0.41-1.75) | 0.83 (0.50-1.37) | 0.89 (0.38-2.08) | 1.07 (0.47-2.41) | 0.88 (0.16-4.80) | 1.08 (0.64-1.84) |
| Umami | W/M vsW/W | 1.17 (0.69-1.97) | 1.26 (0.80-1.96) | 0.94 (0.54-1.64) | 1.56 (0.96-2.53) | 1.29 (0.78-2.11) | 1.50 (0.76-2.95) | 0.42 (0.26-0.68)* |
| | M/M vsW/W | 2.57 (1.43-4.61)* | 1.98 (0.87-4.50) | 2.31 (1.29-4.15)* | 2.92 (1.17-7.33) | 4.78 (1.86-12.30)* | 1.00 (0.18-5.70) | 0.43 (0.24-0.78) |

Bold: nominally significant associations; *significant also after FDR correction, including rare haplotypes (not shown); °all other haplotypes as reference; † for genotype analysis, heterozygotes or homozygotes for variant alleles (M) vs homozygotes for wild-type allele (W, ref); for haplotype analysis, subjects with 1 (w/M) or two (M/M) copies of a specific haplotypes versus subjects with the most frequent haplotype (TTC). Analyses adjusted for age, sex, country.

Supplementary Table 2. Associations between Neuromedin U genotypes and taste thresholds in the IDEFICS children population.

| Taste | Genotype or haplotype contrast† | Genotypes (beta±SE, <i>p</i>) | | | Haplotypes (beta±SE, <i>p</i>) | | | |
|--------|---------------------------------|-----------------------------------|-------------------|-------------------|------------------------------------|-------------------|--------------------|-------------------|
| | | rs6827359 (C) | rs12500837 (C) | rs9999653 (T) | H2 (CCT) | H4 (CTT) | H8 (TTT) | H7 (TTC)° |
| Sweet | W/M vsW/W | -0.35±1.11 (0.75) | -0.08±0.99 (0.93) | -1.43±1.17 (0.22) | -1.01±1.05 (0.34) | -0.83±1.02 (0.41) | -1.79±1.32 (0.18) | 0.69±0.99 (0.49) |
| | M/M vsW/W | -0.68±1.30 (0.60) | 0.03±1.87 (0.99) | -1.83±1.29 (0.16) | -0.28±2.09 (0.89) | -1.46±2.06 (0.48) | -1.97±1.99 (0.32) | 1.48±1.33 (0.26) |
| Salt | W/M vsW/W | 1.12±0.72 (0.12) | 1.24±0.64 (0.054) | 0.24±0.77 (0.75) | 0.68±0.69 (0.32) | -0.10±0.70 (0.88) | -1.51±0.82 (0.07) | 0.70±0.67 (0.29) |
| | M/M vsW/W | 0.67±0.84 (0.43) | 0.04±1.22 (0.97) | -0.34±0.84 (0.69) | 0.00±1.24 (1.00) | -0.41±1.30 (0.75) | -3.25±1.68 (0.053) | 0.02±0.86 (0.98) |
| Bitter | W/M vsW/W | -0.01±0.05 (0.85) | 0.03±0.04 (0.53) | -0.01±0.05 (0.84) | 0.01±0.05 (0.80) | -0.02±0.05 (0.59) | 0.01±0.06 (0.89) | -0.01±0.05 (0.90) |
| | M/M vsW/W | -0.04±0.06 (0.50) | 0.01±0.08 (0.89) | -0.04±0.06 (0.53) | -0.03±0.09 (0.75) | -0.07±0.08 (0.37) | 0.27±0.18 (0.13) | 0.00±0.06 (0.95) |
| Umami | W/M vsW/W | -0.50±0.32 (0.12) | 0.14±0.29 (0.61) | -0.43±0.34 (0.20) | 0.01±0.31 (0.97) | -0.32±0.31 (0.30) | -0.09±0.46 (0.85) | -0.35±0.30 (0.25) |
| | M/M vsW/W | -0.13±0.38 (0.74) | 0.19±0.54 (0.73) | -0.29±0.37 (0.43) | 0.00±0.57 (0.99) | -0.34±0.49 (0.49) | -0.22±1.17 (0.85) | 0.28±0.39 (0.47) |

° All other haplotypes as reference; † for genotype analysis, heterozygotes or homozygotes for variant alleles (M) vs homozygotes for wild-type allele (W, ref); for haplotype analysis, subjects with 1 (w/M) or two (M/M) copies of a specific haplotypes versus subjects with the most frequent haplotype (TTC). Analyses adjusted for age, sex, country. Lower detection threshold values indicate higher sensitivity for a specific tastant.